



Original article

Multiple antibiotic resistance (MAR) index of some bacteria isolated from “JIKO” a herbal drink sold in some parts of Kaduna metropolis

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ABSTRACT

The presence of antibiotics resistance bacteria associated with herbal drinks commonly consumed has been a major public health concern. This study aims at determining Multiple Antibiotic Resistance (MAR) Index from “JIKO” a herbal drink consumed in some parts of Kaduna metropolis. Twenty-six bacteria were isolated from JIKO obtained from Kakuri and Ungwan-Tanko using spread plate method. The isolated bacteria were screened against fifteen (15) antibiotics belonging to five different classes, namely, quinolones (ciprofloxacin, perfloxacin, sparfloxacin, ofloxacin), penicillin (ampiclox, amoxicillin, augmentin), cephalosporin (ceftazidine, cefuroxime), aminoglycosides (streptomycin, gentamycin, septrin), microlides and others (chloramphenicol, erythromycin, nalidixic acid). The antibiotic susceptibility testing of the bacteria revealed that 19 out of the 26 bacteria recorded varying degrees of resistance. The MARI values ranged between 0.1-0.9. The results identified some MAR bacteria of public health concern.

Keywords: Multiple antibiotics resistant index, susceptibility, herbal, jiko, health

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INTRODUCTION

Resistant to antibiotics and its consequence of antibiotic treatment in persons is soaring, leading to serious public health issues of worldwide significance and having an immediate effect in combating infections caused by bacterial (1). Antibiotics intake is a great influence in development, discretion and spread antimicrobial resistant microbes in both animal and human therapeutics (2). This has impelled humans in taking costly second or third-generation antibiotics, persistent hospital admission which then results to expensive medical/or hospital cost, escalated death rate and birth rate and severe global economic meltdown (3,4).

Over dose of antibiotics in persons and animals for treatment has steered up antibiotics resistant strains dissemination into the environment (5,6). In Nigeria, various kinds of research have shown sudden rise in the occurrence of gastrointestinal bacterial resistance for the past fifteen years (7). Resistance bacterial Infections/or diseases are widespread and some of the pathogens causing these infections have developed resistance to different types of antibiotics used against them.

Antibiotic resistance can originate from gene mutations or by horizontal transfer of resistance markers between phylogenetically diverse bacteria (8,9). Bacteria that showed resistant to antibiotic incur their antibiotic desensitized conflict genes. For instance, these genes could code for “efflux” pumps that expel antibiotics from cells, these genes probably give way to enzymes that reduce antibiotics standard, or that

synthetically modify and stimulate the antibiotics.

Multiple antibiotic resistance (MAR) index is designed to be a suitable means to evaluate danger/hazard. It evaluates the overall number of bacteria showing antibiotic resistance and the subsequent danger zone in constant susceptibility screening (10). Thenmozhi *et al.*, (11) stated that MARI values higher than 0.2 indicates high risk sources of contamination with often use of antibiotics while values of less than or equal to 0.2 indicates sources from where antibiotics are seldomly used. Hence, this study was carried out to determining MARI from “JIKO” a herbal drink consumed in some parts of Kaduna metropolis by subjecting the isolates to antibiotic study.

MATERIALS AND METHODS

A total of 30 samples of JIKO was collected from both sites (Kakuri and Ungwan-Tanko market) into sterile bottles and transported to the laboratory of Biological Sciences Department, Nigerian Defence Academy in an ice-box container for bacterial isolation. The bacterial isolates were identified and characterized using conventional methods and the isolates were confirmed using 16S rRNA gene.

Antibiotic susceptibility testing

The bacterial isolates were subjected to antibiotic susceptibility screening using the method initiated by Bauer *et al.*, (12) after standardizing the bacterial isolates to 0.5 McFarland standards. The Antibiotic susceptibility testing of the isolates was carried out using Kirby – Bauer disk diffusion method and was evenly spread on the entire surface of Muller-Hinton agar and then allowed to dry for 5 minutes.

Antibiotics disc was now placed on the surface of the agar and it was incubated at 37°C for 24 hours. Any growth with less than 12 mm in diameter zone around the disc was considered indicative of drug resistance to the bacterial growth according to the guidelines of the Clinical and Laboratory Standard Institute (13). The following discs of antibiotics were used ciprofloxacin 10 µg, perfloxacin 10 µg, Sparfloxacin 10 µg, ofloxacin 10 µg, ampiclox 30 µg, augmentin 25 µg, streptomycin 30 µg, septrin 30 µg, gentamycin 10 µg, erythromycin 10 µg, amoxicillin 30 µg, chloranphenicol 10 µg, cefuroxime, ceftazidime 10 µg, nalidixic acid 30 µg.

Identification of Multidrug Resistance (MDR) bacteria

The number of antibiotic each bacterium was resistant to in the disc diffusion test was recorded as multidrug resistance bacteria. Multidrug resistance (MDR) was recorded as resistant to two or more antibiotics tested

Determination of Multiple Antibiotic Resistance (MAR) Index

Multiple antibiotic resistance (MARI) index was determined as a/b, “a” is the number of antibiotics to which the particular isolate was resistant and “b” is the total number of antibiotics to which the isolate was exposed (14).

RESULTS

Antibiotic susceptibility testing of the bacterial isolates from both sites (n=26)

Bacterial isolates were classified as being susceptible, intermediate resistant and resistant to the antibiotics depending on the inhibition zone diameters when compared to the guidelines of the Clinical and Laboratory Standard Institute. Table 1 and Table 2 reveals the Antibiotics susceptibility testing of gram negative and gram positive bacteria isolates. 19 amongs the 26 bacterial isolates that were isolated from the drink, 5 bacteria from Kakuri and 14 from Ungwan-Tanko reveals varying degrees of resistance to the antibiotics they were tested against as depicted below:

Table 1 Antibiotic susceptibility testing of bacteria isolates from Kakuri (n=11)

Bacterial isolates	ANTIBIOTICS														
	CPX	PEF	SP	OFX	APX	AM	AU	CFU	CEF	S	CN	SXT	CH	E	RD
<i>B. cereus</i> 1	I	I	R	I	R	I	I	S	S	I	I	R	S	S	I
<i>B. cereus</i> 1 a	S	S	S	S	S	R	S	I	I	S	S	S	S	S	S
<i>B. cereus</i> 1b	R	S	S	S	S	S	I	R	I	S	S	I	R	S	I
<i>B. cereus</i> 02	R	S	S	S	S	S	S	S	S	S	S	R	R	S	R
<i>B. cereus</i> 03	R	R	R	S	S	R	S	R	R	R	R	R	R	R	S
<i>B. cereus</i> 04	S	S	S	S	R	S	R	S	S	S	I	I	S	S	S
<i>B. cereus</i> 04a	S	S	S	S	I	S	I	S	I	S	S	S	S	S	S
<i>B. cereus</i> 04b	R	S	S	S	I	I	S	S	S	S	S	S	S	S	S
<i>B. cereus</i> 04c	S	S	R	I	S	S	S	S	S	S	S	S	S	S	S
<i>B. cereus</i> 04c2	S	S	S	I	R	S	S	S	S	S	S	S	S	I	S

*B. cereus*04c3 S I I S I R S S S S S S S S S

KEY: KT: Kakuri, CPX (Ciprofloxacin), PEF (Perfloxacin), SP (Sparfloxacin), OFX (Ofloxacin), APX (Ampiclox), AM (Amoxicillin), AU (Augmentin), CEF (Ceftazidine), CFU (Cefuroxime), S (Streptomycin), CN (Gentamycin), SXT (Septrin), CH (Chloramphenicol), E (Erythromycin), RD (Nalidixic acid), S (Sensitive), R (Resistance), I (Intermediate).

Table 2 Antibiotic susceptibility testing of bacteria isolates from Unguwan -Tanko (n=15)

Bacterial isolates	ANTIBIOTICS														
	CPX	PEF	SP	OFX	APX	AM	AU	CFU	CEF	S	CN	SXT	CH	E	RD
<i>B. cereus</i> 01d	R	S	S	S	I	S	I	S	S	S	S	S	S	S	S
<i>S. plymuthica</i>	R	I	S	R	R	R	R	R	S	S	R	S	R	S	S
<i>B. cereus</i> 2a	S	I	R	I	R	I	S	S	S	R	I	S	S	I	S
<i>B. cereus</i> 2b	R	R	R	R	R	R	R	R	I	R	R	R	R	R	R
<i>B. cereus</i> 2c	R	I	I	I	R	I	S	R	S	S	S	S	S	S	S
<i>B. cereus</i> 2d	R	R	S	I	R	S	S	S	S	S	S	S	S	S	S
<i>B. cereus</i> 2e	R	S	R	I	R	S	S	S	S	S	S	S	S	I	S
<i>B. cereus</i> 2f	S	I	R	S	I	S	S	S	S	I	S	S	R	S	S
<i>B. cereus</i> 2g	R	I	R	I	S	I	S	S	S	R	S	S	S	S	S
<i>B. cereus</i> 2h	R	R	S	S	I	S	I	I	S	S	S	S	S	S	S
<i>B. cereus</i> 2i	R	S	S	S	R	I	S	S	S	S	S	S	S	S	S
<i>B. cereus</i> 3a	S	R	S	I	S	S	S	S	S	S	I	S	S	R	S
<i>B. cereus</i> 3c	S	R	S	I	R	S	S	S	S	I	S	S	S	R	S
<i>B. cereus</i> 3e	R	S	S	I	I	S	R	S	S	I	S	S	S	S	S
<i>B. cereus</i> 3g	S	S	S	I	R	S	S	S	S	R	S	S	S	I	S

KEY: UT: Uguwan-Tanko, CPX (Ciprofloxacin), PEF (Perfloxacin), SP (Sparfloxacin), OF(Ofloxacin), LEV (Levofloxacin), APX (Ampiclox), AM (Amoxicillin), AU (Augmentin), CEF (Ceftazidine), CFU (Cefuroxime), S (Streptomycin), CN (Gentamycin), SXT (Septrin), CH(Chloramphenicol), E (Erythromycin), RD (Nalidixic acid), S (Sensitive), R (Resistance), I (Intermediate)

Multiple antibiotics resistant patterns of bacteria isolates from both sites (n=26)

Multiple antibiotics resistant patterns of the bacterial isolates obtained from JIKO from the two sampling sites and their

percentage of resistance. The percentage of resistance of the bacterial isolates from Kakuri JIKO drink showed 1(13.3%), 2(20%), 1(26.7%) and 1(73.3%) values while the percentage of resistance of the bacterial isolates from Ungwan-Tanko JIKO drink showed 5(13.3%), 6(20%), 1(53.3%) and 1(93.3%) values as depicted in Table 3

Table 3: Multiple antibiotics resistant patterns of bacteria isolates and their percentage of resistance from both sites (n=5 and n=14)

Bacterial isolates	ANTIBIOTICS								
	2	3	4	5	6	7	8	9	10≥
KK (n=5)	1(13.3%)	2(20%)	1(26%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	1(73.3%)
UT (n=14)	5(13.3%)	6(20%)	0(0%)	0(0%)	0(0%)	0(0%)	1(53.3%)	0(0%)	1(93.3%)

KEY: 5 isolates from Kakuri and 14 from Ungwan-Tanko were tested to be Multiple antibiotic resistant. n : no of isolates, K: Kakuri, UT: Ungwan-Tanko

Multiple Antibiotic Resistant (MAR) Index from both sites

The bacterial isolates from Kakuri showed MARI values ranged between 0.1-0.7 while Ungwan-Tanko JIKO drink was 0.1-0.9.

The MARI values 0.3-0.9 indicates existence from high risk contaminated sources with indiscriminate use of antibiotics while MARI values of less than or equal to 0.2 reflects bacteria from sources with less antibiotics usage

Table 4: Multiple Antibiotic Resistant (MAR) Index of bacterial isolates from Kakuri (n=5)

Bacteria isolates	No of antibiotic resistant	No of antibiotic exposed	MARI
<i>B. cereus</i> 1	3	15	0.2
<i>B. cereus</i> 1b	3	15	0.2
<i>B. cereus</i> 02	4	15	0.3
<i>B. cereus</i> 03	11	15	0.7
<i>B. cereus</i> 04	2	15	0.1

Key: KK: Kakuri, MARI: Multiple Antibiotic Resistant (MAR) Index.

Table

5: Multiple Antibiotic Resistant Index (MAR) Index of bacteria isolates from Ungwan-Tanko (n=14)

Bacterial isolates	NO of antibiotic resistant	NO of antibiotic exposed	MARI
<i>S. plymuthica</i>	8	15	0.5
<i>B. cereus</i> 2a	3	15	0.2
<i>B. cereus</i> 2b	14	15	0.9
<i>B. cereus</i> 2c	3	15	0.2
<i>B. cereus</i> 2d	3	15	0.2
<i>B. cereus</i> 2e	3	15	0.2
<i>B. cereus</i> 2f	2	15	0.1
<i>B. cereus</i> 2g	3	15	0.2
<i>B. cereus</i> 2h	2	15	0.1
<i>B. cereus</i> 2i	2	15	0.1
<i>B. cereus</i> 3a	2	15	0.1
<i>B. cereus</i> 3c	3	15	0.2
<i>B. cereus</i> 3e	2	15	0.1
<i>B. cereus</i> 3g	2	15	0.1

Key: UT: Ungwan-Tanko, MARI: Multiple Antibiotic Resistant (MAR) Index.

DISCUSSION

Antibiotics are powerful drugs that are generally harmless and very helpful in combating disease, but when abused antibiotics can actually cause harm. The pathogenic bacteria isolated from these herbal drinks showed wide resistance among the isolates. The high level of *Bacillus cereus* resistance to tested antibiotic is in conformity with the report of (15). *Bacillus cereus* recorded highest multidrug resistance with three (3) isolates showing varying degrees of MAR pattern. In particular one *Bacillus cereus* isolate recorded the highest MAR pattern showing resistant to 14 antibiotics namely: ampiclox, amoxicillin, augmentin, cefuroxime, ciprofloxacin, chloramphenicol, ofloxacin, perfloxacin, sparfloxacin, streptomycin, septrin,

erythromycin, nalidixic acid and gentimycin having 93.3% resistant. This result corresponds with the work of (16) whose antibiotic susceptibility resistance patterns of *Bacillus spp* isolated from herbal medicinal products showed resistance to ciprofloxacin, tetracycline, norfloxacin, augmentin, ceftriaxone, gentamicin, and cotrimoxazole.

Bacillus cereus was the most dominant in the herbal preparation because they are widely distributed in the soil, dust, air and because they are resistant to environmental harmful factors.

Serratia plymuthica recorded with MAR pattern showing resistant to 8 antibiotics (53.3%): ciprofloxacin, perfloxacin, ofloxacin, ampiclox, amoxicillin, augmentin,

cefuroxime and gentamycin. This report is in union with the report of (17) in which *S. plymuthica* from human infection showed sensitive to ciprofloxacin, amoxicillin-clavulanic acid, gentamicin, and cefotaxime.

Multiple Antibiotic Resistant (MARI) Index of isolates from both sites was determined. *Bacillus cereus* recorded MARI values between 0.3-0.9 while *Serratia plymuthica* was 0.5. Hence, reflecting that the isolate *Bacillus cereus* and *Serratia plymuthica* might have originated from contaminated sources that have been obtained from multiple harvest sites, soil and environment or reflecting indiscriminate use of antibiotics. The result is in agreement with the work of (18) whose MARI result of *Bacillus spp* from liquid herbal products ranged between 0.4 – 0.6. Thus, all isolates from this study having MARI values greater than 0.2 and above perhaps be misuse of antibiotics or could be due to expression of resistance genes present in the bacteria. Also, it may be as a result of genes channelled from other bacteria species through horizontal gene transfer. High resistance to multiple antibiotics may present high mortality and can create an opportunity for epidemic outbreaks which are tough to contain. Conclusively, this study shows that the herbal drinks are contaminated with pathogenic bacteria. Therefore, it is mandatory both the consumers and producers of herbal drinks adhere to quality and safety rules.

Recommendation

From our findings, it is recommended that further study should be done to identify the genes responsible for resistance or virulence in these organisms to ensure

curative and preventive measures are been adhere to.

Conflict of Interests

Authors declare that there is no conflict of interest regarding the publication of this paper.

REFERENCES

1. Mather, A.E., Mathews, L., Mellor, D.J., Reeve, R., Denwood, and M.J., Boerlin, P. (2012). An ecological approach to assessing the epidemiology of antimicrobial resistance in animal and human populations“, *Proceedings of the Royal Society*, B 279: 1630–1639
2. Tollefson, L. and W.T. Flynn, (2002). Impact of antimicrobial resistance on regulatory policies in veterinary medicine: Status report. *American Association of Pharmaceutical scientist*, 4: 150-159
3. Rosenberger L.H, Hranjec T, and Politano A.D (2011). Effective cohorting and “ Super iosolation in a single intensive care unit in response to an outbreak of diverse Multidrug Resistant organism” *Surgical Infections*, 12 (5): 345-350
4. Morales E, Cots F, Sala M (2012). “Hospital costs of nosocomial multidrug resistant *Pseudomonas aeruginosa* acquisition” *BMC Health Services Research*, 1:122
5. Akhter, A, Imran M, and Akhter F. (2014). Determination or MAR patterns and indexing among metal tolerant B-lactamase producing *E.*

- coli*. *African journal of Microbiology Research*, 8 (7): 619-627
6. Thenmozhi S, Rjeswari P, Suresh Kumar T, Saipritanga V, Kalpana M (2014). Multi-drug Resistance patterns of Biofilm Forming *Aeromonas hydrophila* from Urine Samples. *International Journey of Pharmaceutical Science and Research*, 5(7): 2908-2918.
 7. Bauer, A.W, Kirby W.M, Sherris J.C, and Turck M. (1996). Antibiotic susceptibility testing by a standardized single disc method. *American Journal of Clinical Pathology*, 45: 493-496.
 8. Clinical and Laboratory Standard Institute (2012). Performance Standards for Antimicrobial Susceptibility testing; twenty second informational supplement. CLSI, Pennsylvania, USA.
 9. Kummerer, K. (2003). Significance of antibiotics in the environment. *Journal of Antimicrobial Chemotherapy*, 52:5-7
 10. Bhavish, S, Param P.S and Mandeep H (2017). Multidrug Resistant *Bacillus cereus* in Fresh Vegetables: A serious Burden to Public Health. *International Journal of Current Microbiology and Applied Sciences*, 6(4): 649-661.
 11. Abdela Y, Yitayih Wondimeneh, Teklay G and Feleke Moges (2016). Occurrence of Potential Bacterial Pathogens and Their Antimicrobial Susceptibility Patterns Isolated from Herbal Medicinal Products Sold in Different Markets of Gondar Town, Northwest Ethiopia. *International Journal of Bacteriology*, Article ID 1959418, 11 pages
 12. Sarika J, Shamma A, Rumpa S, Iqbal R and Kaur (2017). *Serratia plymuthica*: A community-acquired uropathogen. *Indian Journal of Medical Sciences*, 69(1):31
 13. Ujam N.T, Oli A.N, Ikegbunam N.M, Adikwu M.U and Esimone C.O (2013). Antimicrobial Resistance Evaluation of Organisms Isolated from Liquid Herbal Products Manufactured and Marketed in South Eastern Nigeria. *British Journal of Pharmaceutical Research*, 3(4): 548-562